



Effect of *Phyllanthus niruri* and *Passiflora foetida* Extracts on the Mortality and Survival rate of the Brine Shrimp *Artemia salina*

Juario Juzavil¹, Mondejar Eddie¹, Nuñeza Olga¹ and Uy Mylene²

¹Department of Biological Sciences, College of Science and Mathematics, Mindanao State University, Iligan Institute of Technology, A. Bonifacio Ave., Tibanga, Iligan City, 9200, PHILIPPINES

²Department of Chemistry, College of Science and Mathematics, Mindanao State University, Iligan Institute of Technology, A. Bonifacio Ave., Tibanga, Iligan City, 9200, PHILIPPINES

Available online at: www.isca.in, www.isca.me

Received 20th May 2014, revised 24th July 2014, accepted 4th September 2014

Abstract

Phyllanthus niruri and *Passiflora foetida* are known to have pharmacological properties. This study tested the lethality effects of the different extract concentrations of *P. niruri* and *P. foetida* on brine shrimps in order to assess the bioactivity of the plants. The sampled plants were taken from selected areas in Mindanao, Philippines. Plant extractions were done using three solvents: water, absolute ethanol and 50:50 ethanol-water. LC50 values were calculated to evaluate the toxicity of the plant extracts. The LC50 values of *P. niruri* extracts were >1000 µg/ml for plant extract with water as solvent, 251.19 µg/ml for plant extract with absolute ethanol as solvent, and 281.84 µg/ml for plant extract with 50:50 ethanol-water solvent. The LC50 values of *P. foetida* with the same solvents were >1000 µg/ml, 749.89 µg/ml and >1000 µg/ml, respectively. These results suggest a low toxicity level of the plant extracts. This implies that the use of these medicinal plants especially in the traditional approach which is through the decoction method has the least risk of danger. The results also suggest that *P. niruri* and *P. foetida* may contain bioactive compounds needing further elucidation and tests.

Keywords: Bioactivity, concentrations, lethality, pharmacological, toxicity.

Introduction

Interest on medicinal plants to ameliorate health care problems is increasing locally and internationally¹. Over the years, advances in drugs and other pharmaceutical products originating from plants are revealed and are introduced². Many developing countries address health concerns by depending on phytomedicine. Most of the people in Africa make use of traditional herbal medicine to meet their health requirements³. In addition, the use of traditional remedies has already been engaged since the Ayurvedic treatment in India ranging from Asia and to Europe⁴.

Passiflora foetida (Linn.) belonging to Passifloraceae family is an herbaceous climber with the auxiliary tendrils and yellowish white hair stems. It is frequently found growing in thin soil. It is native in tropical America and is wild in several parts of India⁵. *Passiflora* species are significant because of their high therapeutic values. Tribes with traditional medicinal knowledge recommended *P. foetida* L. as a good source of high value pharmaceutical plant⁶.

P. foetida is a common weed found in farmlands where it could invade monocrops⁷ and could affect the yield of the farmers. Nevertheless, its invasive ability in agricultural aspects is paralleled with its benefit as this weed is tested to have useful features. Because of its substantial effect, it may be necessary to yield additional medicines to cure or ease illnesses. *P. foetida*

leaf extract is reported to possess anti-inflammatory effects and analgesic activity⁸; anti-histamine⁹; anti-depressant effects¹⁰ and estrogenic activity¹¹. The fruit extract is reported to have anti-inflammatory effects¹² and the whole plant possesses immunomodulatory effects¹³. Traditional medicinal practitioners of Chittor District of India use it against asthma and various neurological disorders¹⁴. In addition to its curative effect in nervous ailments, it is also used to treat problems on arthritis¹⁵. Other species of *Passiflora* is used as sedative, antidote for anxiety and hypertension¹⁶.

Phyllanthus had been widely used as ethnomedicinal agent around the world¹⁷. Recently, there is a rising consciousness of reviewing and developing the pharmacological and biochemical assets of this plant because of its active mechanism. This invites further research for health care applications¹⁸. Six different species of *Phyllanthus*, dominated by *Phyllanthus amarus*, are sold for their homeopathic causes in the market of southern India¹⁹. Many are already reported about the positive effects of *Phyllanthus*, used as a therapeutic treatment against hepatic issues^{20,21}, renal complications²² and can also be antioxidant and reactive oxygen species scavenger^{23,24}.

Phyllanthus niruri is one of the species of *Phyllanthus*. It is an herbaceous plant with small, green, short-petiole and bitter-taste leaves; has thin and glaucous under surface; unisexual, monoecious and minute flowers in the axil of the leaves²⁵. Some *Phyllanthus* species in some countries like Thailand propagate

together in open places, wastelands and secondary forest²⁶. In a study conducted using RAPD polymorphism, *Phyllanthus* species were distinguished from each other. *Phyllanthus urinaria*, *P. debilis* and *Phyllanthus embilica* were found grouped into one major group in dendrogram based on the RAPD polymorphism yet the *P. urinaria*, *P. debilis* and *P. amarus* have more resemblances in vegetative morphology²⁷.

Phyllanthus belongs to the family Euphorbiaceae- family of flowering plants. In the Philippines, there are more than 40 *Phyllanthus* species. The common names for the genus used in this study are "sampasampalukan, surusampalok, talikod, or taltalikod, San Pedro, malakirum-kirum, turutalikod" and other tags depending on the local or cultural terms. These were found as weeds growing throughout the Philippines at low and medium altitudes²⁸.

Passiflora foetida, like *P. niruri*, is also a weed with agricultural significance. It is usually grown to reduce soil erosion or to impede the growth of *Imperata cylindrica* grass ("kogon") in a coconut plantation²⁹.

Phyllanthus, tagged as a "wonder plant" because of its numerous beneficial effects³⁰, has been widely studied for its species classification, phylogeny and morphology of its flower³¹; phytochemical observation³²; phytochemical screening, anti-inflammatory and cytotoxic activities^{33,34}, efficacy of its extracts in induced hepatitis³⁵, and phytochemical and pharmacological properties³⁶. There are limited studies on the toxicity of *P. niruri*³⁷. Further investigation of this medicinal plant is needed in the Philippines to supplement the existing lethality evaluation done on medicinal plants in the country³⁸. Toxicity test is essential to evaluate the lethal implications of the medicinal plant to be used³, and may suggest the plant's supplementary bioactivity³⁹. This further draws attention to any biochemically significant compounds found in the plant⁴⁰. The application of brine shrimp lethality bioassay is mostly general because it is functional, low-cost and fast assessment requiring only ample sample size and requirement⁴¹.

This study used brine shrimp lethality test to investigate the toxicity level of *P. niruri* and *P. foetida*, to know if these plant species have possible hazard when employed as therapeutic agents and to test the bioactivity of the plant extracts which could be of high importance to further pharmacological studies on the plants.

Material and Methods

Plant Collection and Identification: The plant samples of *P. niruri* and *P. foetida* were collected from Mt. Matutum of South Cotabato; Kalubihon, Tibanga and Brgy. Abuno of Iligan City, Philippines in the month of August 2013. The collected samples were properly labelled and identified. Plant identification was based on taxonomic keys and published works⁴².

Crude extracts preparation: One to two kilograms (kg) of

fresh samples of leaves and stems was properly washed with tap water and rinsed with distilled water. The rinsed samples were air-dried for one week or until when the samples were already crispy enough upon crumpling. The dried samples of each plant were pulverized using a sterile electric blender. The powder was weighed, divided into two equal parts and stored in glass containers. One part was saturated with enough absolute ethanol and the other part was soaked in 50:50 ethanol - water for three days (72 hours). Each solution was filtered using Whatman filter paper and collected in a glass container. Adequate amount of the filtered solution was then concentrated in a rotary evaporator to achieve the crude plant extract. After extracting, the 50:50 ethanol-water extract was freeze dried to eliminate the excess water.

Plant decoction preparation: About 450grams (g) of fresh and clean samples of the plants were cut into pieces and boiled in about 900 ml distilled water corresponding to a 1:2 ratio of water and sample for decoction for five minutes. The mixture was then filtered. The solvent was then freeze dried to remove traces of water, then cooled and stored in glass containers until needed for the lethality testing.

Toxicity Assay: Brine shrimps hatching: Brine shrimp eggs were hatched in sterile seawater. A rectangular glass aquarium was divided into two compartments with holes on the partition. The eggs were placed into the first compartment (covered) where the brine shrimp eggs hatched while the second compartment was illuminated. Mature nauplii (larvae) swam towards this illuminated compartment. Eggs hatched after 24 hours incubation at room temperature (25-29°C) with constant exposure to light. They were then collected by glass capillary from the lighted side and added into each test tube.

Brine Shrimp Lethality Test: Thirty milligrams (ml) of the extracts from each fraction were dissolved with a sufficient amount of solvent to obtain 10,000 ppm stock solution. In triplicate, serial dilutions (1000 µg/ml, 500 µg/ml, 100 µg/ml, and 10 µg/ml) were made in each test tube and dissolved in dimethyl sulfoxide (DMSO). Ten nauplii and seawater were added to each well obtaining a 5ml volume of each tube. The test tubes were examined and the number of dead (non-motile) nauplii in each test tube was counted after 24 hours.

Statistical Analysis: Brine shrimps were exposed to four different extract concentrations to determine the relative toxicity of the extracts⁵¹. The relationship between the concentration of the extracts and mortality of the brine shrimps was shown by plotting the concentration log (x-axis) versus mortality (y-axis). LC50 is the dose that resulted to 50% mortality of the brine shrimps.

Results and Discussion

Results showed that there is an increasing mortality rate of the brine shrimps with increasing concentration of the extracts. This suggests a direct proportional relation between concentration

and mortality. Table-1 shows that the absolute ethanol extract of *P. niruri* has the most potent activity with LC50 value of 251.19 µg/ml. The *P. foetida* extracts had the lowest toxicity. Extracts with LC50 values of 1.0 - 10µg/ml are treated as toxic while values > 100µg/ml are considered non-toxic⁴³. *P. niruri* and *P. foetida* results failed to show highly toxic levels of extracts.

Table-1
Chronic LC50 values of *Phyllanthus niruri* and *Passiflora foetida* extracts against the Brine Shrimp *Artemia salina*

| Plant | Extract | Chronic LC50 (24 hours), µg/ml |
|---------------------------|---------------------|--------------------------------|
| <i>Phyllanthus niruri</i> | Decoction/water | >1000 |
| | Absolute ethanol | 251.19 |
| | 50:50 ethanol-water | 281.84 |
| <i>Passiflora foetida</i> | Decoction/ water | >1000 |
| | Absolute ethanol | 749.89 |
| | 50:50 ethanol-water | >1000 |

The results of this study showed that at increased concentrations, *P. niruri* and *P. foetida* have variable levels of toxicity. The mortality of the brine shrimps administered with different concentrations of the *P. niruri* and *P. foetida* extracts may be attributed to the possible constituents or compounds present in them which have beneficial effects as therapeutic agents parallel to an investigation evaluating medicinal plants⁴³.

The leaves and stems of the sampled plant extracts used here presented low toxicity or non-toxicity as shown in the LC50 values which are higher than 100µg/ml. This means that the plant could not convey harm upon critical exposure because of its minor or low toxicity, particularly the water extracts (decoction). This suggests that the decoction, which is commonly practiced in traditional medicine, has no distinct toxic effect in acute administration⁴³. The low toxicity effect using the brine shrimp lethality test was also reported on four Indonesian plants²⁴. The study may have resulted into twice the value of LC₅₀ compared to the present study but both outcomes denote a low cytotoxic properties present in the *P. niruri* extract. This is because most toxic

extracts showed an LC₅₀ of less than 1µg/ml but not exceeding 100µg/ml⁴³. This minute level of harmful effect is parallel with another study, wherein the whole plant extract was found to be non-toxic and not genotoxic with the acute administration at 300mg/kg in rats³.

Greater shrimp death indicates high noxiousness at 50% or less concentration and, for the extract to be considered slightly toxic,

it needs to cause 50-70% cell death in higher dilutions. In toxicology, pharmacological compounds found even at higher concentrations (non-toxic dose) draw possibilities of useful and functional toxic compounds⁴⁴.

The LC50 values of *Phyllanthus acidus* were 3.12 µg/ml, 12.5 µg/ml and 70% mortality above 50 µg/ml and considered highly toxic. High lethality indicated by the brine shrimp lethality test suggests the presence of cytotoxic elements and high bioactivity of the plant extract. The extract of *P. amarus* (synonymous to *P. niruri*⁴⁵) is found to have tannins and phenols having healing significance³². Aside from the limited literatures regarding the toxicity of *P. niruri*, change in environmental factors could also affect the chemical properties of the plant. Thus, more tests are recommended about its chemical properties after some time, to confirm prevailing data³⁷.

This study showed the lethality effect of 749.89 µg/ml of *P. foetida* leaf and stem extracts implying its low toxicity. The necessity of elevated concentration value may be the same with the 200mg/kg and 300mg/kg doses of its leaf extracts as a painkiller to the ache caused by Eddys hot plate^{46,47} and almost the same effect with the study on the activities of *P. foetida* L8. This trivial level of lethality relates to its chronic toxicity ineffectiveness when tested in mice⁴⁸. The report revealed that 20g/kg dose administered exhibited neither any abnormality to the tested group, nor significant changes in the body weight, food intake and relative organ weight compared to the control group. Alteration in haematological parameters and biochemical parameters both range at doses 800 to 1600mg/kg which are still covered in the normal range. In a study, the ethanol leaf extract of the plant did not bring about any death or abnormality to tested mice even at the highest dose of 200mg/kg after three days⁸. A study also showed that acetone and ethanol extracts of *P. foetida* leaves have anti-microbial property⁴⁹. It validated the plant's curative activity in higher concentrations and traditional treatment of infections affected by microbes or bacteria.

The threshold of toxicity could be considered to have significant biological activity, proposing strong injurious compounds which warrant further investigation⁵⁰. Brine shrimp assay has been considered as an examination for antimicrobial, antitumor, anti-malarial, antifungal and insecticidal activities⁵¹. The present results merit further investigation of *P. niruri* and *P. foetida* plants to verify their medicinal potentials.

Conclusion

P. niruri and *P. foetida* extracts showed no toxicity as indicated by the need to have a higher concentration of the extracts to trigger mortality on the brine shrimps. This points out that usage of the plant extracts as a therapeutic agent poses no high risk of harm/ toxicity upon acute administration especially when prepared in a traditional way which is the decoction method. However, the death of some of the nauplii also suggests that *P.*

niruri and *P. foetida* may have compounds that need further research given that some literatures have shown some of their antioxidant activities and other biochemical properties.

Acknowledgment

We acknowledge the DOST ASTHRD for the financial support.

References

1. Krishnaraju A.V., Tayi V.N., Roa T.V.N., Sundararaju D., Vanisree M., Tsay H-S. and Subbaraju G.V., Assessment of Bioactivity of Indian Medicinal Plants Using Brine Shrimp (*Artemia salina*) Lethality Assay, *International Journal of Applied Science and Engineering*, **3(2)**, 125-134, Downloaded on October 15, 2013 from http://s3.amazonaws.com/academia.edu.documents/31068189/IJASE__6_%2807-003%29- libre.pdf? AWSAccessKeyId=AKIAJ56TQJRTWSMTNPEA & Expires=1398354525&Signature=VyTrFZo3dHVVHsr IU 8osxeO5Nuhk%3D, (2005)
2. Habib M.D.R, Sayeed M.A., Rahman M.D.M., Hasan M.D.R. and Saha A., In Vitro: Evaluation Of Cytotoxic, Antibacterial, Antioxidant And Phytochemical Screening Of Petroleum Ether Extract Of *Phyllanthus Acidus*, *IJPSR*, **2(4)**, 875-881, Downloaded on October 15, 2013 from <http://www.ijpsr.com/v2i4/19%20Vol.%202, Issue%204,IJPSR,2011,Paper%209.pdf>, (2011)
3. Asare G.A., Bugyei K., Sittie A., Yahaya E.S., Gyan B., Adjei S., Addo P., Wiredu E. K, Adjei D.N. and Nyarko A.K., Genotoxicity, cytotoxicity and toxicological evaluation of whole plant extracts of the medicinal plant *Phyllanthus niruri* (Phyllanthaceae), *Genet. Mol. Res.*, **11(1)**, 100-11, Downloaded on October 10, 2013 from <http://www.funpecrp.com.br/gmr/year2012/vol11-1/pdf/gmr1315.pdf>, (2012)
4. Thyagarajan S.P., Jayaram S., Gopalakrishnan V., Hari R., Jeyakumar P. and Sripathi MS., Herbal medicines for liver diseases in India, *Journal of Gastroenterology and Hepatology*, **17**, S370-S37, Downloaded on October 10, 2013 from http://www.researchgate.net/profile/Sripathi_Sureban/publication/10998933_Herbal_medicines_for_liver_diseases_in_India/file/d912f50d6652328051.pdf (2002)
5. The wealth of India, Raw materials, CSIR, New Delhi, **7**, 278 Downloaded on October 10, 2013 from <http://www.niscair.res.in/activitiesandservices/products/wealth-of-indiaFolder2010.pdf>, (1966)
6. Patil A.S. and Paikrao H.M., Bioassay Guided Phytometabolites Extraction for Screening of Potent Antimicrobials in *Passiflora Foetida* L., *Journal of Applied Pharmaceutical Science*, **2(9)**, 137-142, Downloaded on October 18, 2013 from http://www.japsonline.com/admin/php/uploads/648_pdf.pdf (2012)
7. Takim, F.O., Olaoye A.O. and Adeyemo J., Survey of *Passiflora foetida* L. and Associated Weed Species on Arable Crops in Ballah, Southern Guinea Savanna Zone Of Nigeria, *Agrosearch*, **12(2)**, 117 – 12 Downloaded on October 18, 2013 from http://www.japsonline.com/admin/php/uploads/648_pdf.pdf (2012)
8. Sasikala V., Saravanan S. and Parimelazhagan T., Analgesic and anti-inflammatory activities of *Passiflora foetida* L., *Asian Pacific Journal of Tropical Medicine*, **4(8)**, 600-603, Downloaded on October 18, 2013 from <http://apjtm.net/admin/picture/UploadFile/20111229164259667.pdf> (2011)
9. Chivapat S., Bunjob M., Shuaoprom A., Bansidhi J., Chavalittumrong P., Rangsriripat A. and Sincharoenpokai P., Chronic toxicity of *Passiflora foetida* L. extract, *International Journal of Applied Research in Natural Product* **4(2)**, 24-31, Downloaded on October 18, 2013 from <http://www.ijarnp.org/index.php/ijarnp/article/download/8/8> (2011)
10. Santosh P., Venugopl R., Nilakash A. S., Kunjibhari S. and Mangala L., Antidepressant Activity of Methanolic Extract of *Passiflora foetida* Leaves In Mice, *International Journal of Pharmacy and Pharmaceutical Sciences*, **3(1)**, 112-115, Downloaded on October 18, 2013 from http://www.researchgate.net/profile/Kunjibhari_Sulakhiya/publication/235721358_ANTIDEPRESSANT_ACTIVITY_OF_METHANOLIC_EXTRACT_OF_PASSIFLORA_FOETIDA_LEAVES_IN_MICROE/file/9fcfd512dfd862e14e.pdf (2011)
11. Michel B.G., Koffib K., Stanislasa Z.O., Alassanec T. and Flaviena T., Oral acute toxicity and estrogenic effects of the extracts of *Passiflora foetida* Linn. (Passifloraceae) leaves in female Wistar albino rats, *Annals of Biological Research*, **3(9)**, 4609-4616, Downloaded on October 18, 2013 from <http://scholarsresearchlibrary.com/ABR-vol3-iss9/ABR-2012-3-9-4609-4616.pdf> (2012)
12. Fernandez J., Noronha M. A. and Fernandes R., Evaluation of Anti-inflammatory activity of stems of *Passiflora foetida* Linn. in rats, *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, **4(2)**, 1236-4, Downloaded on October 18, 2013 from [www.rjpbcs.com/pdf/2013_4\(2\)/\[134\].pdf](http://www.rjpbcs.com/pdf/2013_4(2)/[134].pdf) (2013)
13. Ranganatha N., Kuppast I.J., Veerashekar T. and Kulkarni S., Assessment of Immunomodulatory Activity of Aerial Parts of *Passiflora foetida* L., *World Journal of Pharmacy and Pharmaceutical Sciences*, **2(3)**, 1176-1186, Downloaded on October 18, 2013 from http://www.wjpps.com/get_file.php?type=article&file=13751778274%20WJPPS%20479.pdf (2013)
14. Krishnaveni A. and Thaakur S.R., Pharmacognostical

- and preliminary phytochemical studies of *Passiflora foetida*, *Ancient Science of Life*, **27(3)**, 19-23, Downloaded on October 18, 2013 from <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3330862/pdf/ASL-27-19.pdf> (2008)
15. Rajan S., Sethuraman M. and Baburaj D.S., Plants From The Traditional Medical System Of The Nilgiri Tribes, *Ancient Science of Life*, **16(4)**, 360-365, Downloaded on October 18, 2013 from <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3331168/pdf/ASL-16-360.pdf> (1997)
16. Ingale A.G. and Hivrale A.U., Pharmacological studies of *Passiflora* sp. and their bioactive compounds, *African Journal of Plant Science*, **4(10)**, 417-426, Downloaded on October 18, 2013 from http://www.researchgate.net/publication/259220704_Pharmacological_studies_of_passiflora_sp_and_their_bioactive_compounds (2010)
17. Onocha P.A. and Ali M.S., Antileishmaniasis, phytotoxicity and cytotoxicity of Nigerian Euphorbiaceae Plants 2, *Phyllanthus amarus* and *Phyllanthus muellerianus* Extracts, *African Scientist*, **11(2)**, 1595-6881 Downloaded on October 9, 2013 from <http://klobex.org/journals/afs/afs11/afs1120110021.pdf> (2010)
18. Calixto J.B., Santos A.R.S., Filho V.C. and Yunes R.A., A review of the plants of the genus *Phyllanthus*: Their chemistry, pharmacology, and therapeutic potential, *Med Res Rev*, **18(4)**, 225-258 Downloaded on October 10, 2013 from [http://onlinelibrary.wiley.com/doi/10.1002/\(SICI\)1098-1128\(199807\)18:4%3C225::AID-MED2%3E3.0.CO;2-X/abstract;jsessionid=2002_F2DE1598BE3FE5C024785FDA26A2.f03t01](http://onlinelibrary.wiley.com/doi/10.1002/(SICI)1098-1128(199807)18:4%3C225::AID-MED2%3E3.0.CO;2-X/abstract;jsessionid=2002_F2DE1598BE3FE5C024785FDA26A2.f03t01) (1998)
19. Srirama R., Senthilkumar U., Sreejayan N., Ravikanth G., Gurumurthy B.R., Shivanna M. B., Sanjappa M., Ganeshaiyah K.N. and Shaanker R.U., Assessing species admixtures in raw drug trade of *Phyllanthus*, a hepatoprotective plant using molecular tools, *Journal of Ethnopharmacology*, **130 (2)**, 208-215 Downloaded on October 10, 2013 from http://www.ecbol.org/docs/Publications/Srirama_2010.pdf (2010)
20. Liu J., Lin H. and McIntosh H., Genus *Phyllanthus* for chronic hepatitis B virus infection: a systematic review, *J Viral Hepat*, **8(5)**, 358-66 Downloaded on October 10, 2013 from <http://www.ncbi.nlm.nih.gov/pubmed/11555193> (2001)
21. Xin-Hua W., Chang-Qing L., Xing-Bo G. and Lin-Chun F., A comparative study of *Phyllanthus amarus* compound and interferon in the treatment of chronic viral hepatitis B, *Southeast Asian J Trop Med Public Health*, **32(1)**, 140-2 Downloaded on October 10, 2013 from http://www.tm.mahidol.ac.th/seameo/2001_32_1/22-2132.pdf (2001)
22. Freitas A.M., Schor N. and Boim M.A., The effect of *Phyllanthus niruri* on urinary inhibitors of calcium oxalate crystallization and other factors associated with renal stone formation, *B. J. U. Int*, **89(9)**, 829-834 Downloaded on October 10, 2013 from <http://www.ncbi.nlm.nih.gov/pubmed/12010223> (2002)
23. Mahdi E.S., Sakeena M.H.F., Muthanna F.M., Ghassan Z.A., Sattar M.A. and Noor A.M., In Vitro Study of Antioxidants Activity Potential of 30% Ethanolic Extracts Derived from Two *Phyllanthus* Species Indigenous to Malaysia, *Malaysian Journal of Pharmaceutical Sciences Suppl.*, **1**, 159 Downloaded on October 10, 2013 from [http://web.usm.my/mjps/MJPS%208%20SUPP%201%202010/MJPS%20ART%202%20\(145-199\).pdf](http://web.usm.my/mjps/MJPS%208%20SUPP%201%202010/MJPS%20ART%202%20(145-199).pdf) (2010)
24. Nurcholis W., Priosoeryanto B.P., Purwakusumah E.D., Katayama T. and Suzuki T., Antioxidant, Cytotoxic Activities and Total Phenolic Content of Four Indonesian Medicinal Plants, *Valensi*, **2(4)**, 501-510 Downloaded on October 10, 2013 from http://www.researchgate.net/profile/Waras_Nurcholis/publication/235657231_Antioxidant_Cytotoxic_Activities_and_Total_Phenolic_Content_of_Four_Indonesian_Medicinal_Plants/file/32bfe5125a7c0d1145.pdf?ev=pub_ext_doc_dl&origin=publication_detail&inViewer=true (2012)
25. Ross I., *Phyllanthus niruri*, *Medicinal Plants of the World*, **1**, 393-403 Downloaded on October 10, 2013 from http://link.springer.com/chapter/10.1007%2F978-1-59259-365-1_22 (2003)
26. Chantaranothai P., *Phyllanthus*. In: Santisuk T. and Larsen K. (eds). Flora of Thailand. Bangkok: Prachachon, **8**, 473-507 (2007)
27. Theerakulpisut P., Kanawapee N., Maensiri D., Bunnag S. and Chantaranothai P., Development of species-specific SCAR markers for identification of three medicinal species of *Phyllanthus*, *Journal of Systematics and Evolution*, **46(4)**, 614-621 Downloaded on October 10, 2013 from <http://www.plantsystematics.com/qikan/manage/wenzhang/aps07123.pdf> (2008)
28. Merrill E. D., An enumeration of Philippine flowering plants, *Bureau of Printing, Manila*, **2**, 391-396 Downloaded on October 10, 2013 from <http://www.biodiversitylibrary.org/ia/enumerationofphi02merr#page/397/mode/1up> (1923)
29. Patil A.S. and Paikrao H.M., Bioassay Guided Phytometabolites Extraction for Screening of Potent Antimicrobials in *Passiflora Foetida* L., *Journal of Applied Pharmaceutical Science*, **2(9)**, 137-142 Downloaded on October 10, 2013 from http://www.japsonline.com/admin/php/uploads/648_pdf.pdf (2012)
30. Nair R.R., and Abraham R.S., Integrating the Science of

- Pharmacology and Bio Informatics *Phyllanthus*"The wonder plant", *Advanced BioTech*, **6(7)**, 28-30 Downloaded on October 10, 2013 from <http://www.advancedbiotech.in/05%20integrating.pdf> (2008)
31. Chen Y-J., Chen S-H., Huang T-C. and Wu M-J. Pollen Morphology of Philippine Species of *Phyllanthus*(*Phyllanthaceae*, *Euphorbiaceae* s.l.), *Blumea*, **54(1-3)**, 47–58 Downloaded on October 10, 2013 from <http://docserver.ingentaconnect.com/deliver/connect/nhn/00065196/v54n1/s12.pdf?expires=1398359555&id=77960269&titleid=75002419&acname=Guest+User&checksum=4F7B0855849766EB3BF0EA EFC97521D6> (2009)
32. Chandrashekar K.S., Joshi A.B., Satyanarayana D. and Pai P., Phytochemical Observation Of Whole Plant of *Phyllanthus debilis* Klein .Ex.Willd, *Ancient Science of Life*, **25 (1)**, 39-41 Downloaded on October 10, 2013 from <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3330894/pdf/ASL-25-39.pdf> (2005)
33. Zimudzi C., Gwenhure L.F., Kunonga N., Kativu S. and Jere J., Phytochemical screening, cytotoxicity and anti-inflammatory activities of the Zimbabwean endemic plant *Phyllanthus serpentinicola* Radcl.-Sm. (*Phyllanthaceae*), *Journal of Applied Pharmaceutical Science*, **2(10)**, 050-053 Downloaded on October 10, 2013 from http://www.japsonline.com/admin/php/uploads/665_pdf.pdf (2012)
34. Chakraborty R., Biplab D., Devanna N., and Sen S., Antiinflammatory, antinociceptive and antioxidant activities of *Phyllanthus acidus* L. extracts, *Asian Pacific Journal of Tropical Biomedicine*, **2(2)**, S953-S961 Downloaded on October 10, 2013 from <http://www.apjtb.com/zz/2012s2/89.pdf> (2012)
35. Samali A., Tarfa F.D., Odeniran O.A., Onanuga C.E. and Adzu B., Evaluation of efficacious activities of aqueous extract of *Phyllanthus niruri* against acetaminophen-induced hepatitis in rats, *Int. J. Biol. Chem. Sci.*, **6(3)**, 920-930 Downloaded on October 10, 2013 from <http://www.ajol.info/index.php/ijbcs/article/viewFile/80639/70879> (2012)
36. Bagalkotkar G., Sagineedu S.R., Saad M.S. and Stanslas J., Phytochemicals from *Phyllanthus niruri* Linn. and their pharmacological properties: a review, *Journal of Pharmacy and Pharmacology*, **58**, 1559–1570 Downloaded on October 10, 2013 from <http://onlinelibrary.wiley.com/store/10.1211/jpp.58.12.0001/asset/jpp.58.12.0001.pdf?v=1&t=hueacs1v&s=e314f08c3512102b02b891544584fe0b9246297f> (2006)
37. Asare G.A., Addo P., Bugyei K., Gyan B., Adjei S., Otu-Nyarko L.S., Wiredu E.K. and Nyarko A., Acute toxicity studies of aqueous leaf extract of *Phyllanthus niruri*, *Interdiscip Toxicol.*, **4(4)** 206–210 Downloaded on October 10, 2013 from http://www.intertox.sav.sk/ITX_pdf/04_04_2011/10102-Volume4_Issue_4-06_paper.pdf (2011)
38. Olowa L.F. and Nuñeza O.M., Brine Shrimp Lethality Assay of the Ethanolic Extracts of Three Selected Species of Medicinal Plants from Iligan City, Philippines, *International Research Journal of Biological Sciences*, **2(11)**, 74-77 Downloaded on October 10, 2013 from <http://www.isca.in/IJBS/Archive/v2/i11/12.ISCA-IRJBS-2013-177.pdf> (2013)
39. Ved C.H., More N.S., Bharate S.S. and Bharate S. B., Cytotoxicity Screening of Selected Indian Medicinal Plants using Brine-Shrimp Lethality Bioassay, *Advances in Natural and Applied Sciences*, **4(3)**, 389-395 Downloaded on October 10, 2013 from <http://www.aensiweb.com/anas/2010/389-395.pdf> (2010)
40. Pisutthanana S., Plianbangchang P., Pisutthanana N., Ruanruaya S. and Muanrit O., Brine Shrimp Lethality Activity of Thai Medicinal Plants in the Family Meliaceae, *Naresuan University Journal*, **12(2)**, 13-18 Downloaded on October 10, 2013 from http://www.nupress.grad.nu.ac.th/journal/index.php/NUJ_ournal/article/download/232/239 (2004)
41. Krishnarajua A.V., Tayi V.N., Raoa, T.V.N., Sundararajua D., Vanisreeb M., Tsayb, H-S. and Subbaraju, G.V., Assessment of Bioactivity of Indian Medicinal Plants Using Brine Shrimp (*Artemia salina*) Lethality Assay, *International Journal of Applied Science and Engineering*, **3(2)**, 125-134 Downloaded on October 10, 2013 from www.cyut.edu.tw/~ijase/2005/IJASE%20%203-2-6.pdf (2005)
42. Madulid D., Plant diversity in the Philippines. In: Biodiversity and terrestrial ecosystems, *Institute of Botany, Academia Sinica Monograph Series No. 14*, 105-109. (1995)
43. Elnour E.A., Abdmageed E.M. A.M., Shyoub M. E. and Mohammed R.R., Evaluation Of Toxicity Of Some Plants Having Traditional Uses In Sudan On Brine Shrimp, *Global J Trad Med Sys*, **2(1)**, 19-23 Downloaded on October 10, 2013 from <http://www.gjtms.info/index.php/gjtms/article/viewFile/23/8> (2013)
44. Golla, U.R., Gajam P.K., Mohammad A.R., Kumar A. and Solomon Sunder Raj S.S., Assessment of Bioactivity of *Desmostachya bipinnata* (L.) Stapf using Brine Shrimp (*Artemia salina*) Lethality Assay, *Pharmacologyonline*, **3**, 982-990 Downloaded on October 10, 2013 from http://www.researchgate.net/profile/Upendar_Golla/publication/234094464_ASS ESSMENT_OF_BIOACTIVITY_OF_Desmostachya_bipinnata_%28L.%29_Stapf_USING_BRINE_SHRIMP_%28ARTEMIA_SALINA%29_LETHALITY_ASSAY/file/79e4150f044f18862f.pdf?ev=pub_ext_doc_dl&origin

- =publication_detail&inViewer=true (2011)
45. Taylor L., Technical Data Report for Chanca Piedra "Stone Breaker" (*Phyllanthus niruri*). In: Herbal Secrets of the Rainforest, 2nd edition, Sage Press, Inc. Downloaded on October 10, 2013 from www.rain-tree.com/chanca-techreport.pdf (2003)
 46. Chan S.K., Bajrang R.D., Shasshidhar S.B. and Jayaram R.M., Evaluation of analgesic activity of hydro alcoholic leaf extract of *Passiflora foetida*, *Indian J Pharmacol. Supplement*, 2(40), 75 Downloaded on October 10, 2013 from <http://www.ijpbs.net/vol-4/issue-1/pharma/31.pdf> (2008)
 47. Patil A. S., Paikrao H. M. and Patil S.R., *Passiflora foetida* Linn: a complete morphological and phytopharmacological review, *Int J Pharm Bio Sci*, 4(1), 285-296 Downloaded on October 10, 2013 from www.ijpbs.net/vol-4/issue-1/pharma/31.pdf (2013)
 48. Chivapat S., Bunjob M., Shuaoprom A., Bansidhi J., Chavalittumrong P., Rangsripipat A. and Sincharoenpokai P., Chronic toxicity of *Passiflora foetida* L. extract, *International Journal of Applied Research in Natural Products*, 4 (2), 24-31 Downloaded on October 10, 2013 from <http://www.ijarnp.org/index.php/ijarnp/article/viewFile/8/8> (2011)
 49. Mohanasundari C., Natarajan D., Srinivasan K., Umamaheswari S. and Ramachandran A., Antibacterial properties of *Passiflora foetida* L. – a common exotic medicinal plant, *African Journal of Biotechnology*, 6(23), 2650-2653 Downloaded on October 10, 2013 from <http://www.ajol.info/index.php/ajb/article/viewFile/58170/46534> (2007)
 50. Reed L.J and Muench H., A simple method of estimating fifty percent endpoints, *The American Journal of Hygiene*, 27, 493-497 Downloaded on October 10, 2013 from aje.oxfordjournals.org/content/27/3/493.full.pdf (1938)
 51. Krishnarajua A.V., Tayi V.N., Raoa T.V.N., Sundararajua D., Vanisreeb M., Tsayb H-S. and Subbaraju G.V., Biological screening of medicinal plants collected from Eastern Ghats of India using *Artemia salina* (Brine Shrimp Test), *Int. J. Appl. Sci. Eng.*, 4(2), 115-125 Downloaded on October 10, 2013 from www.cyut.edu.tw/~ijase/2006/4.2-08-008-2.pdf (2006)